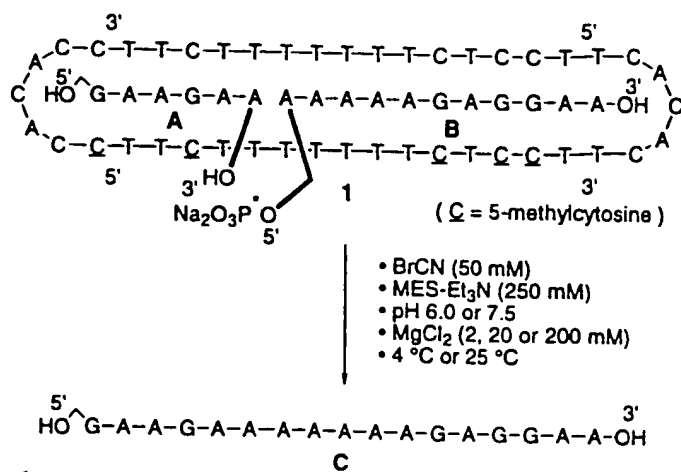




Figure 1. Top: triplex resulting from homopurine ligation fragments bound to pyrimidine bases of the circular DNA template (**R** represents backbone of ligating fragments). Note the use of 5-methylcytidine on the Hoogsteen side of the circular template. Bottom: ribbon graphic of a ligation reaction of two ODNs directed by a circular DNA template.



Scheme 1.

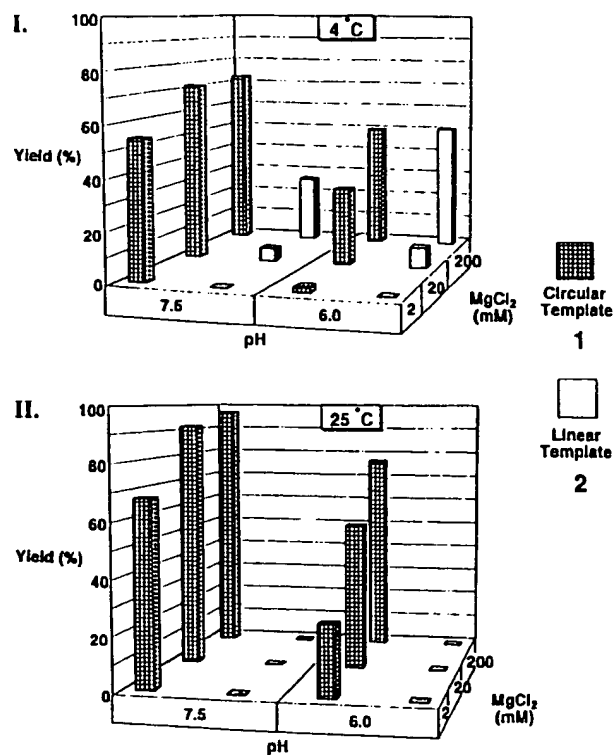


Figure 2. 3-D bar graphs showing the yield (%) of ligation product C. Graph I shows ligation results at 4°C, pH 7.5 and 6.0 with MgCl₂ concentrations of 2, 20 and 200 mM. Graph II shows the same ligation reactions run at 25°C. Data for these graphs was obtained at a reaction time of 3 h. All reactions were reproduced at least twice to afford a % yield error of ± 3 .

Figure 3

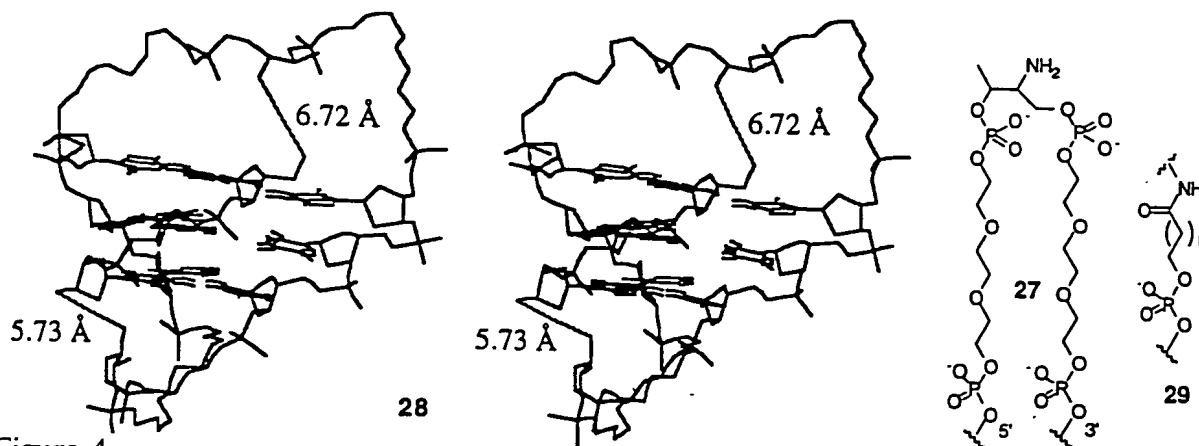
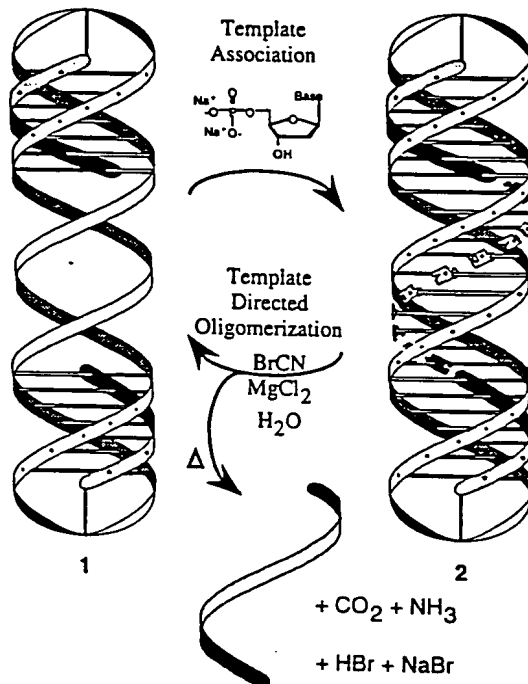
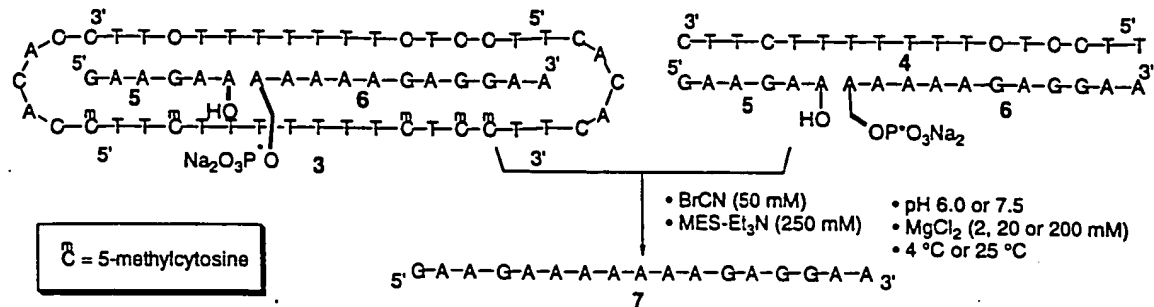


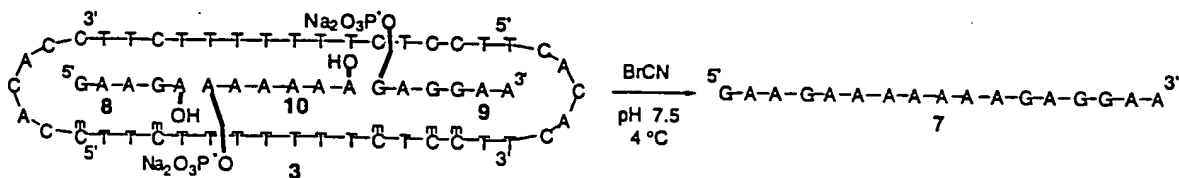
Figure 4 . Stereoview of the energy minimized³⁶ circular DNA template (28) with looped linker 27 attached at both ends of the truncated triplex. Based on structure 28, the initial template-primer linker for investigation will be 29 ($n = 2$). Based on an MM2 minimized structure, 29 will span a distance up to 7 \AA .

Scheme 2.

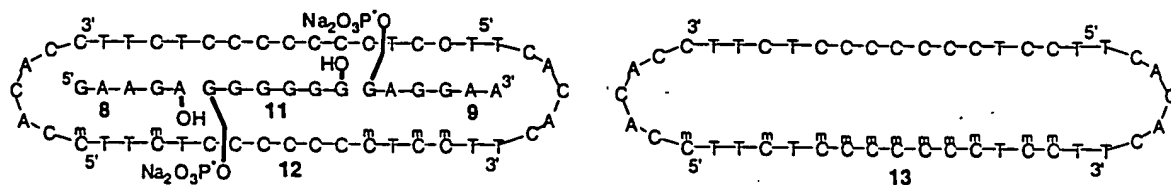


\overline{C} = 5-methylcytosine

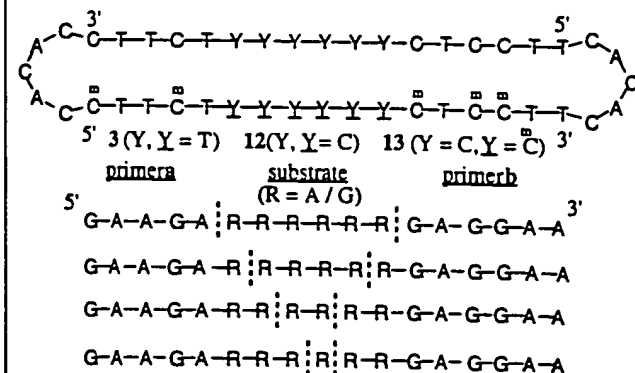
Scheme 3.



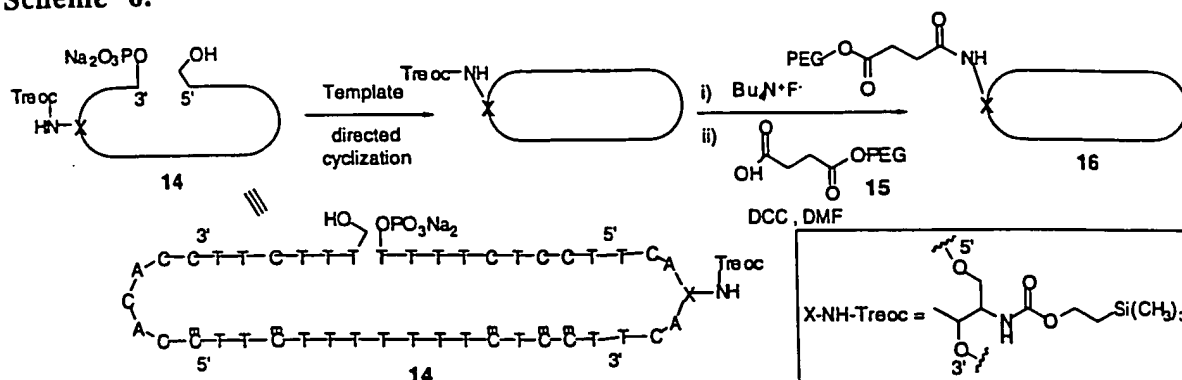
Scheme 4.



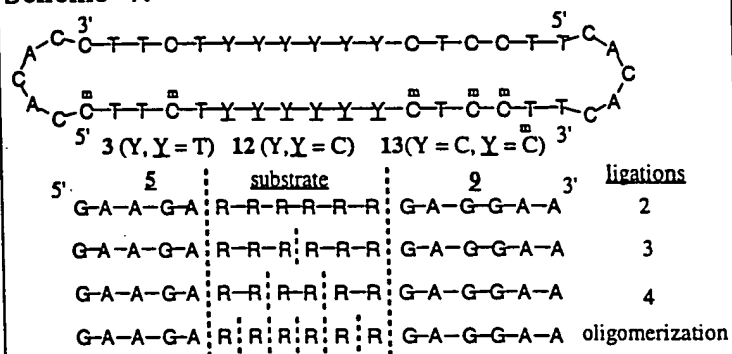
Scheme 5.



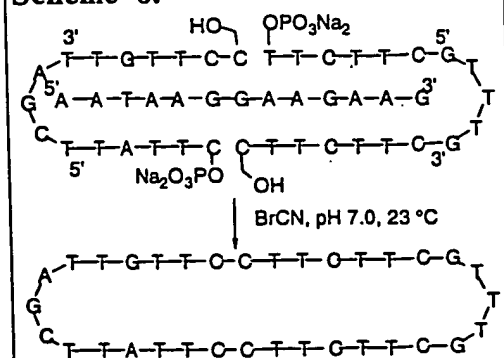
Scheme 6.



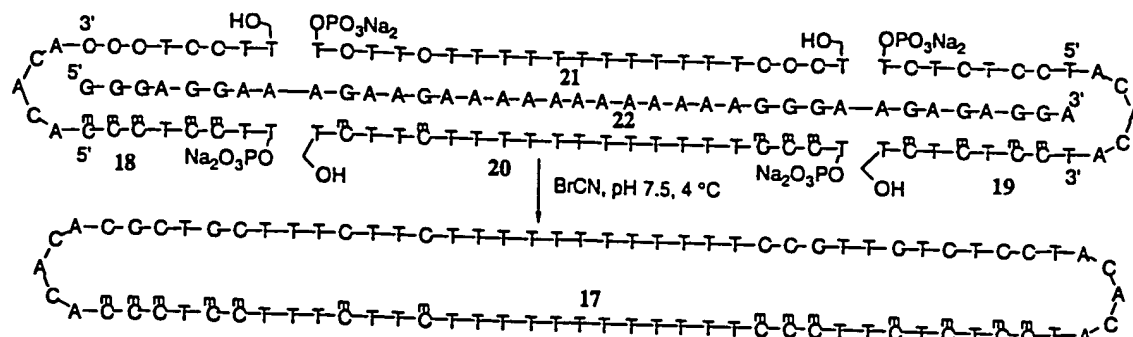
Scheme 7.



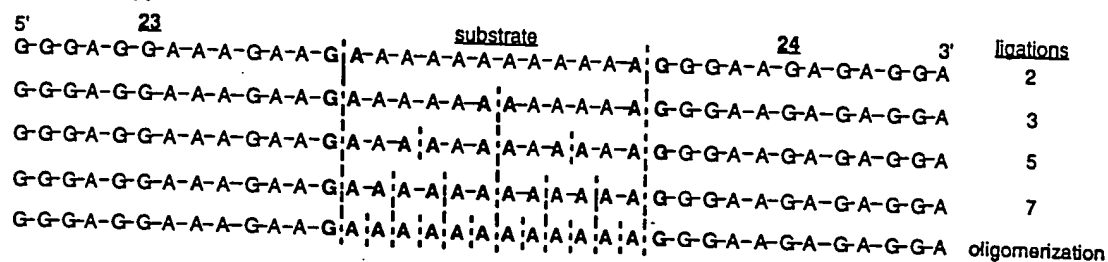
Scheme 8.



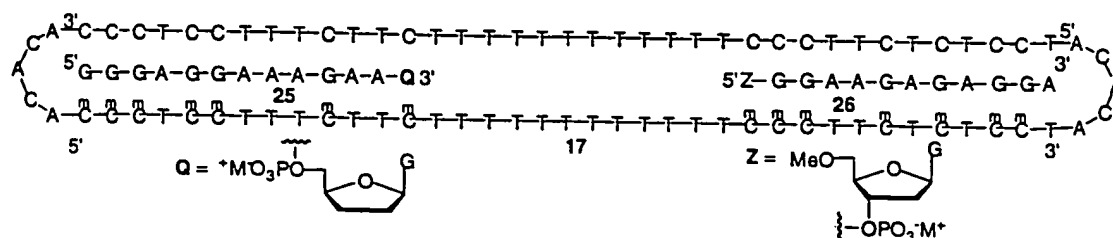
Scheme 9.



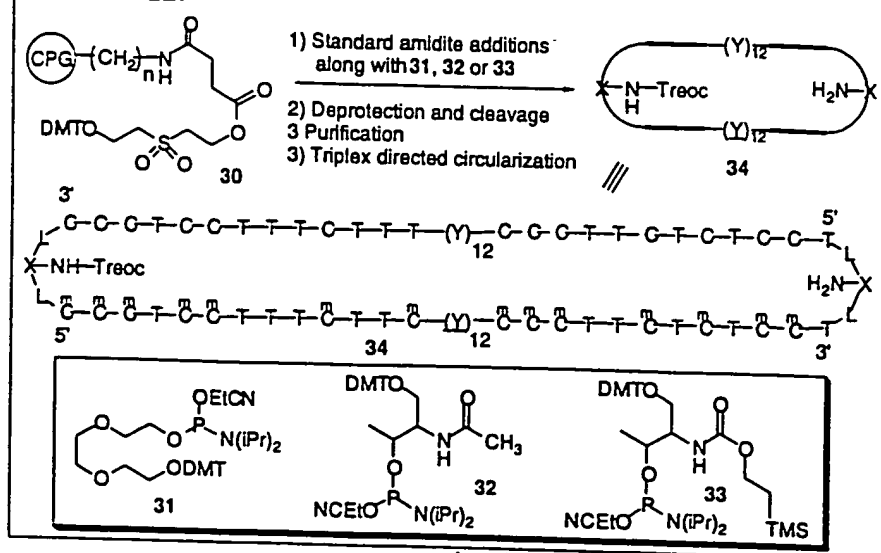
Scheme 10.



Scheme 11.



Scheme 12.



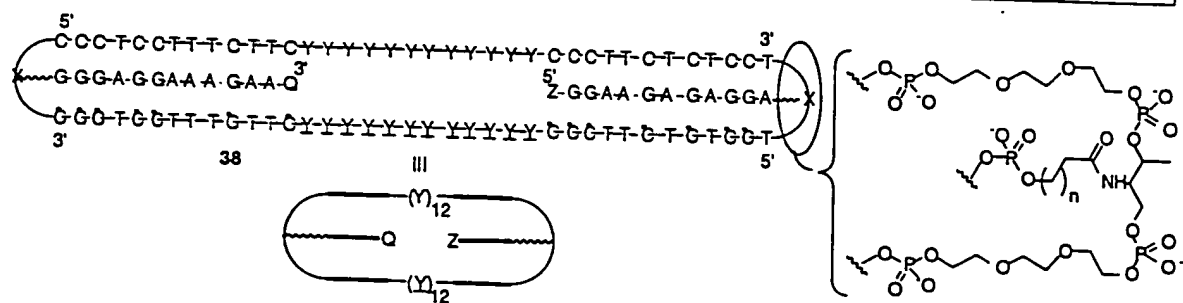
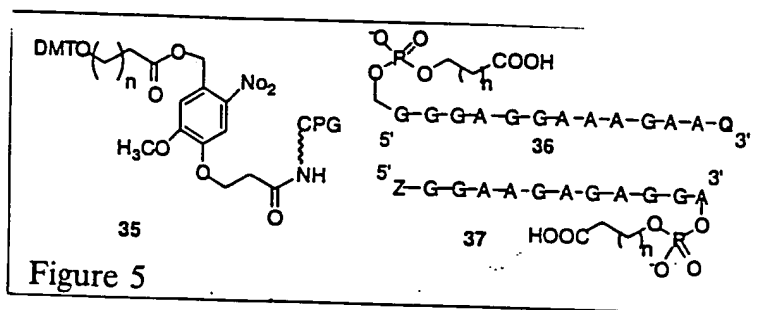
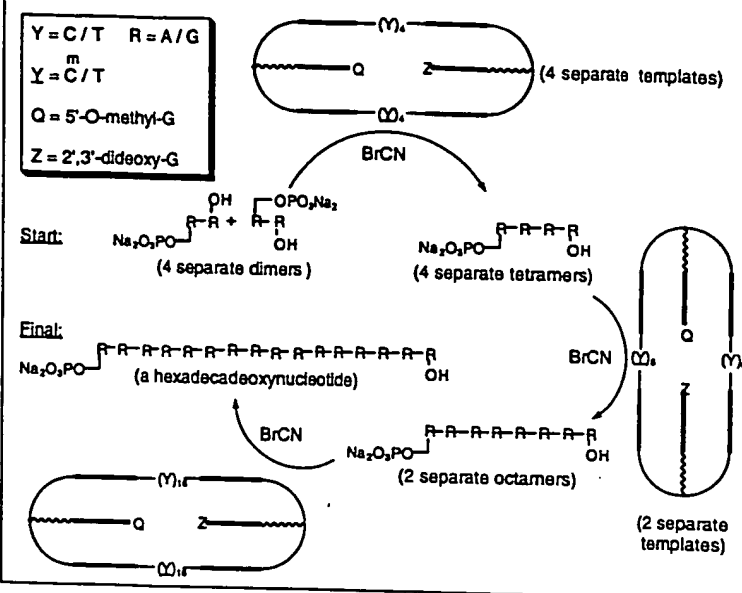
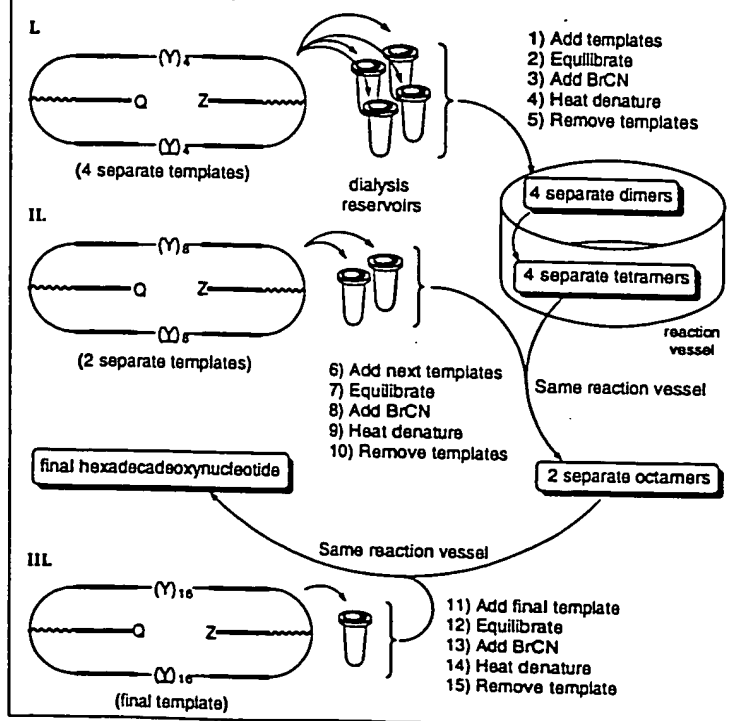


Figure 6 . Primer attached circular DNA template. See Scheme 11 for Q and Z designation.

Scheme 13.



Scheme 14.



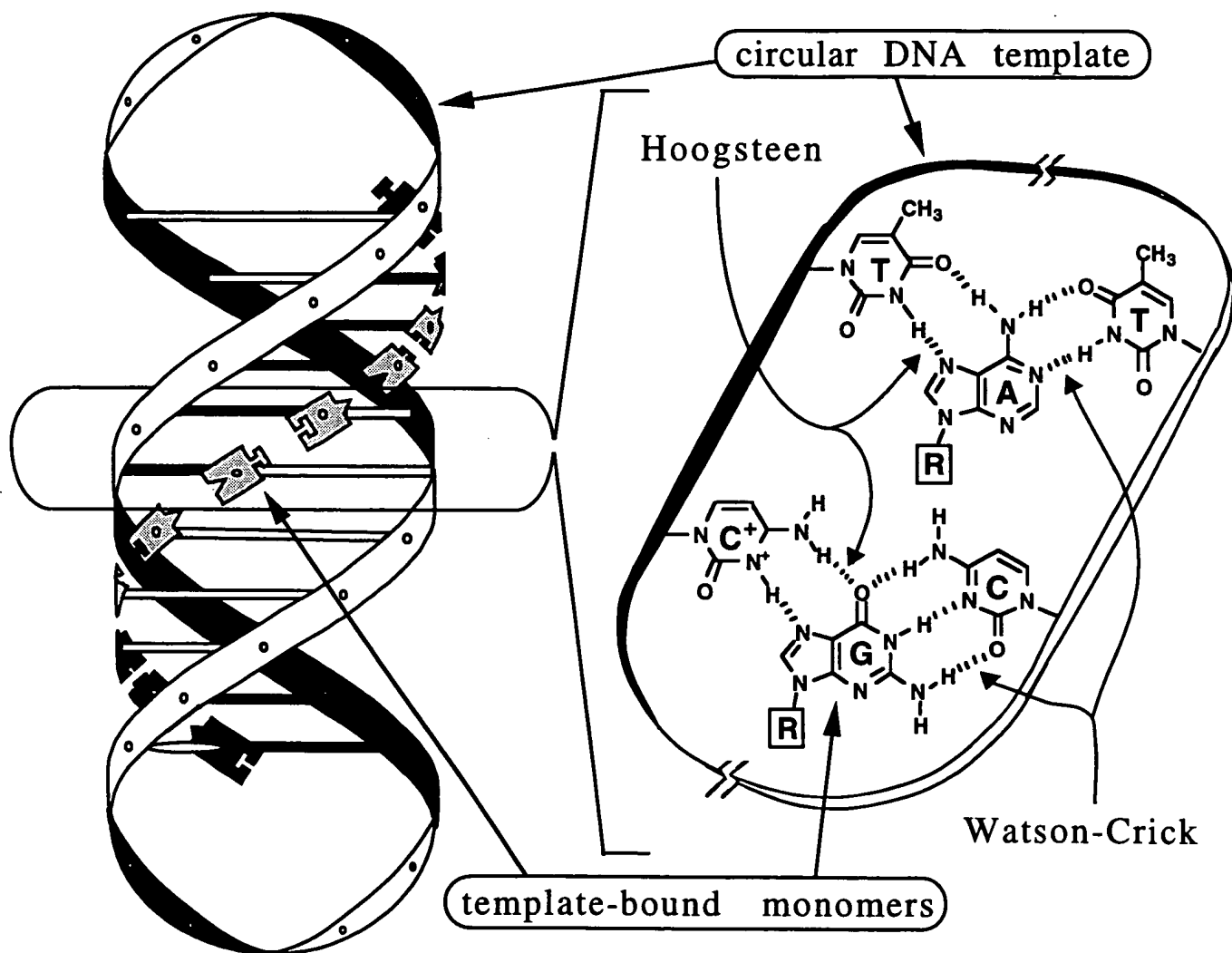


FIG. 7 Ribbon graphic of a circular DNA template with bound monomer nucleobase derivatives and illustration of the template-substrate triplets (**R** represents reacting substrate for oligomerization).

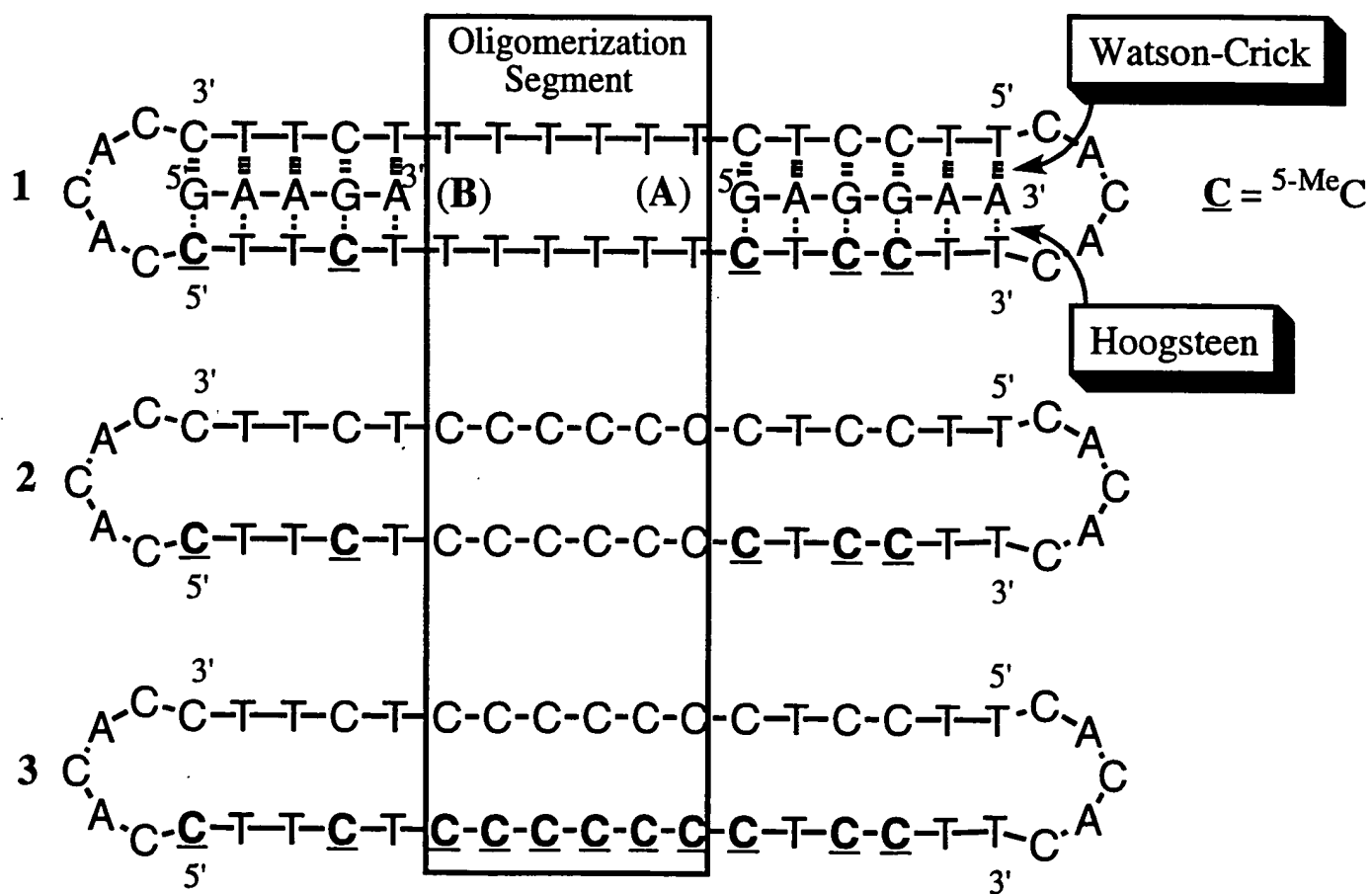
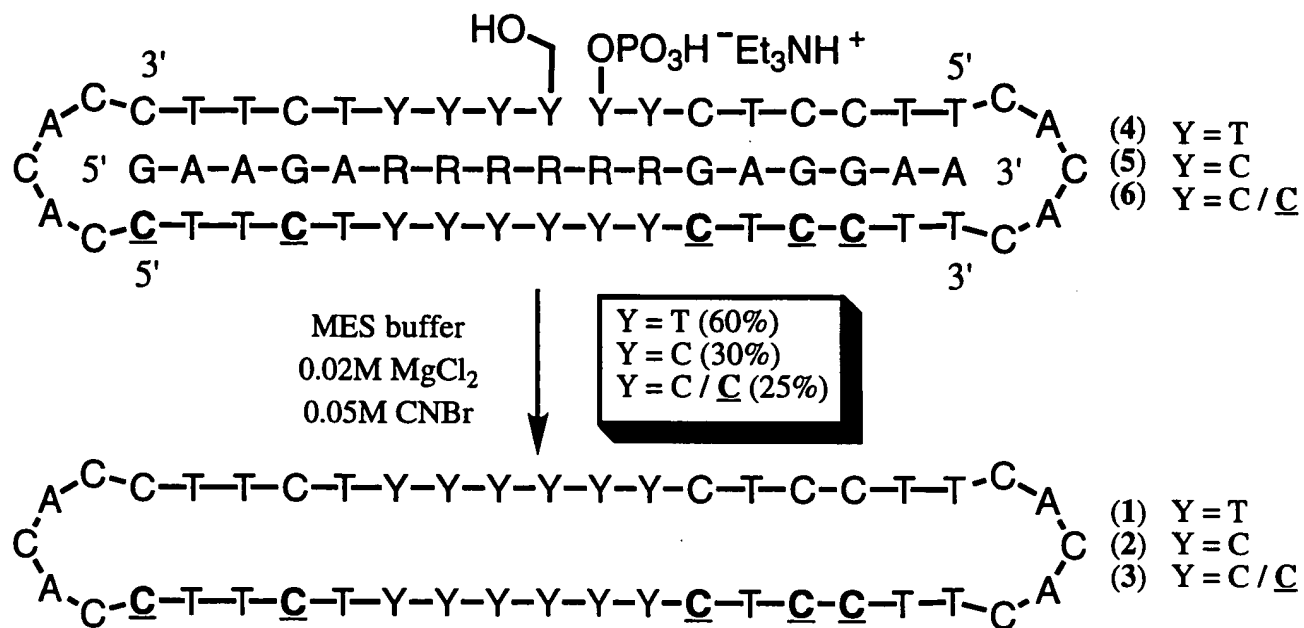


FIG. 8 Three Circular DNA templates (1, 2, and 3) and their respective primers (A and B) for directed ligation and oligomerization experiments. The C designates 5-methyl-cytidine.

Scheme 15



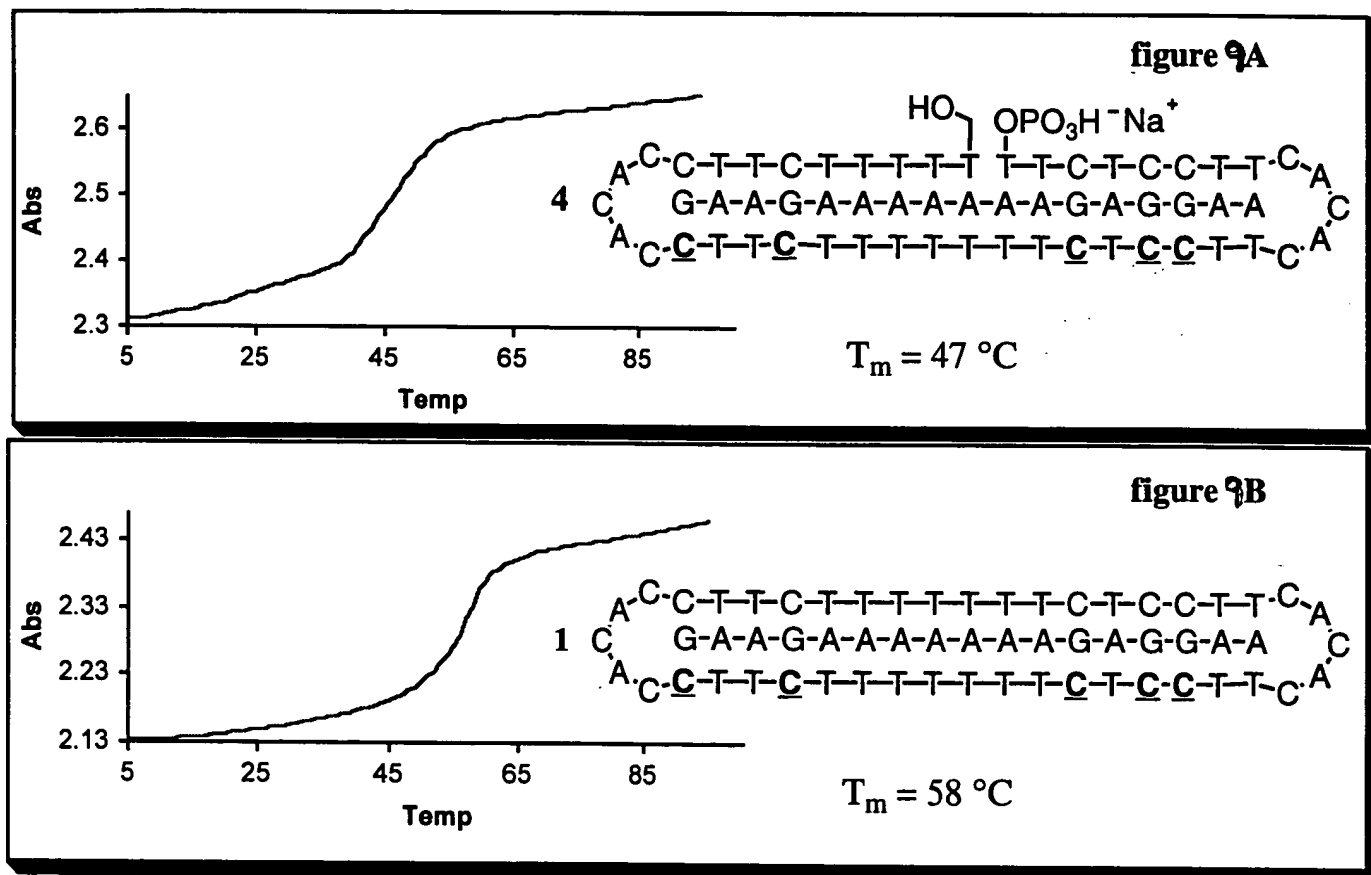


FIG. 9 Melting analysis of 4 with the purine-rich oligonucleotide (3A) and circularized template 1 with the same oligonucleotide (3B).

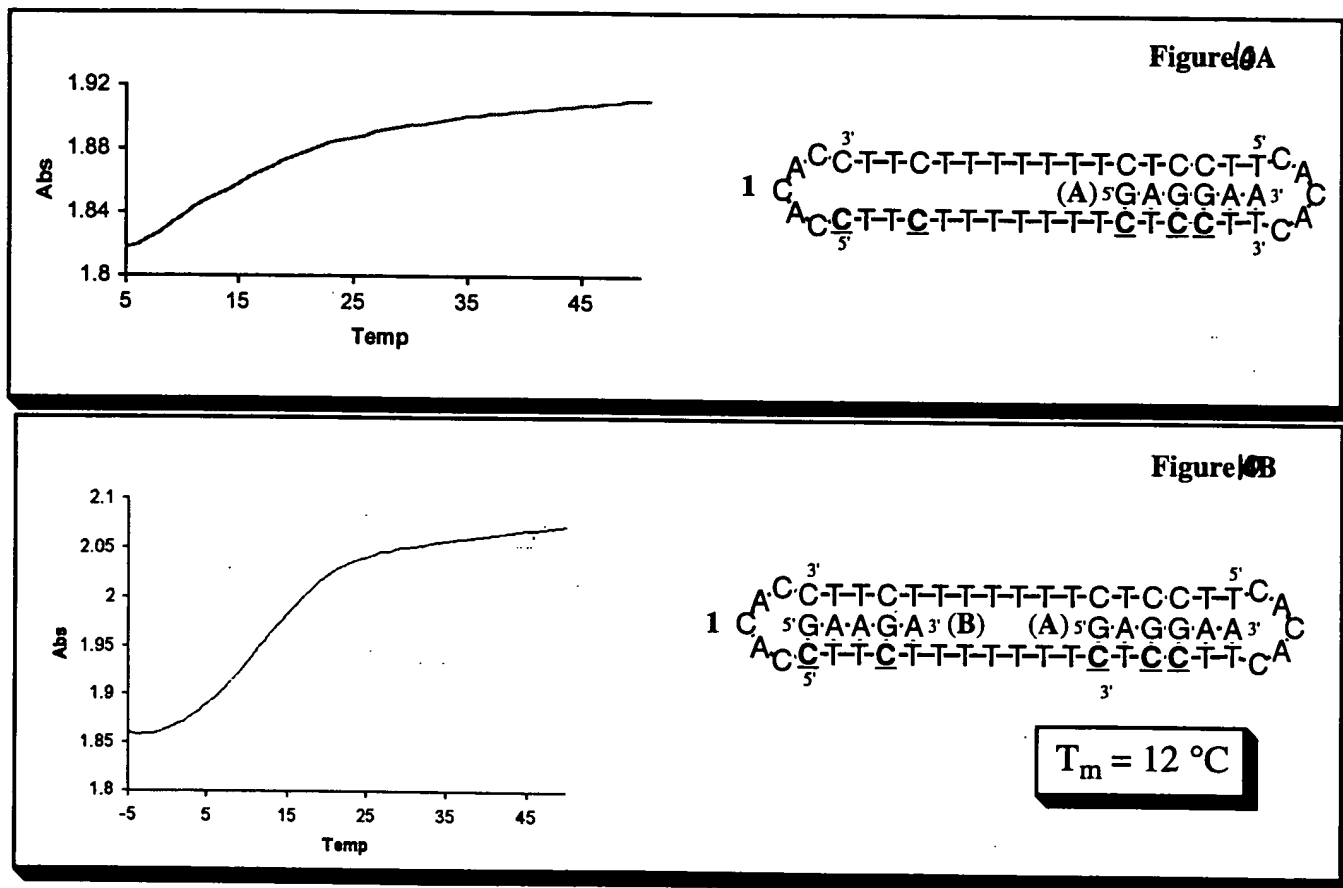


FIG. 10 Melting analysis of circular template 1 with primer A (fig 0A) and primers A and B (fig 0B).

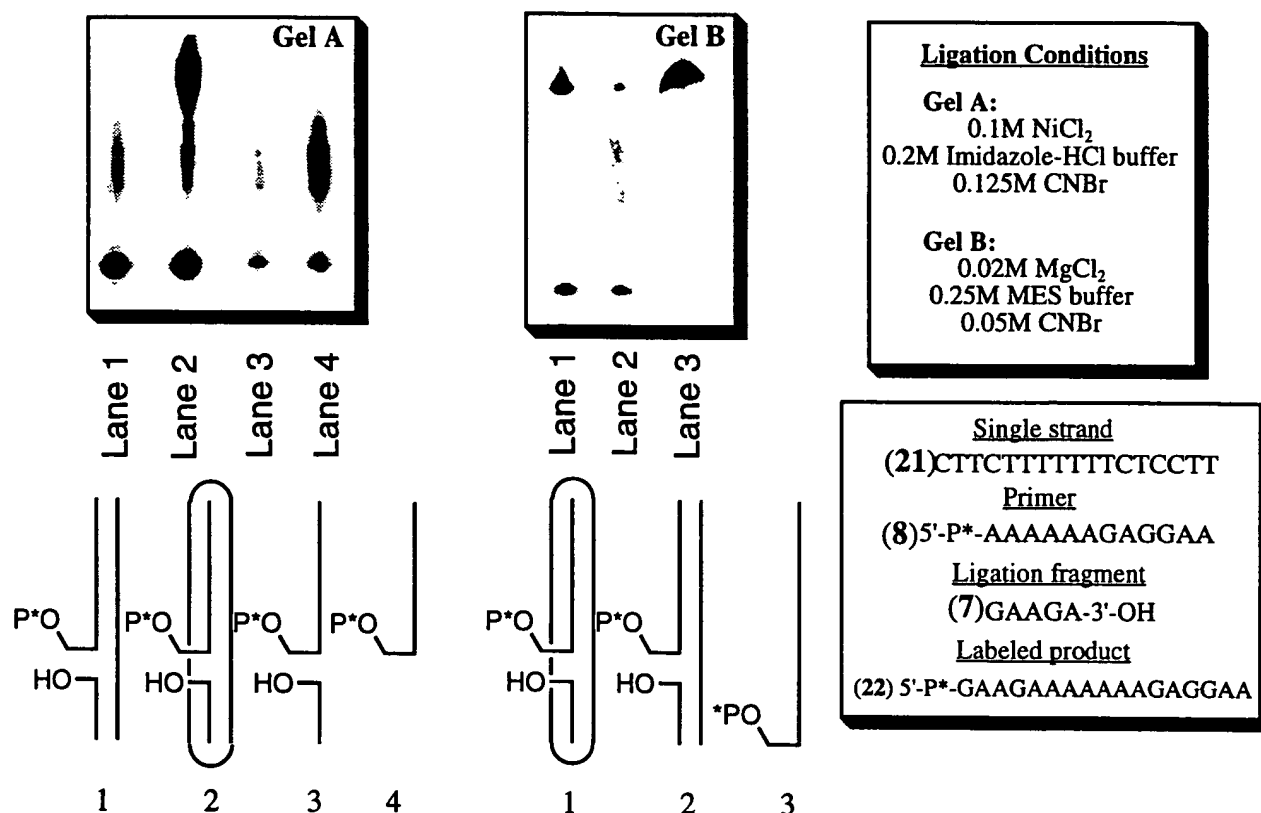
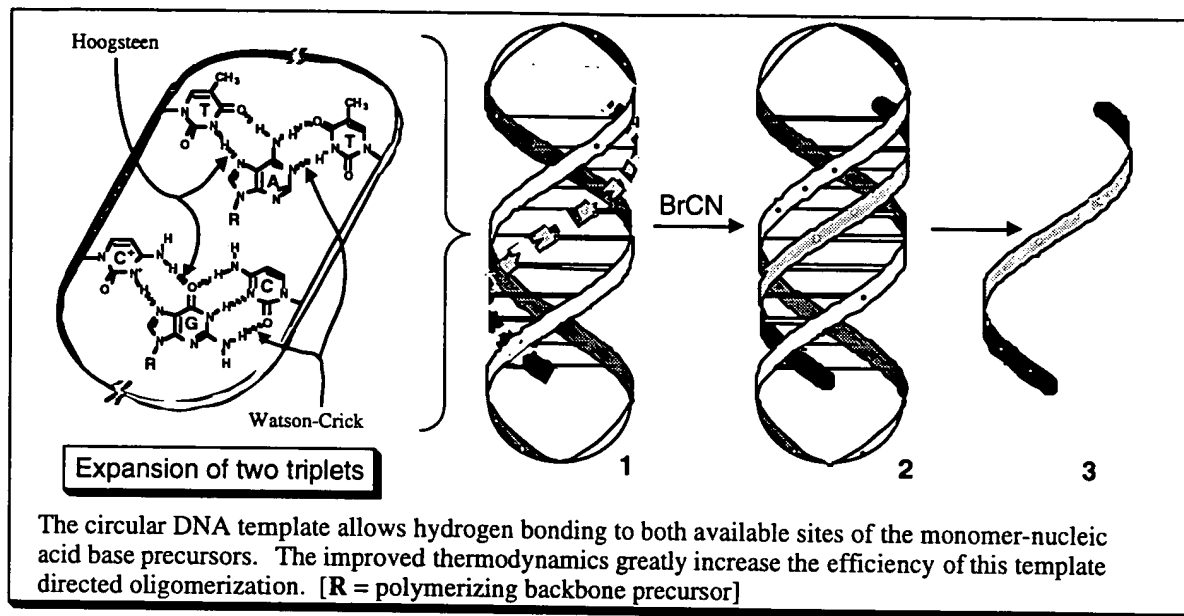
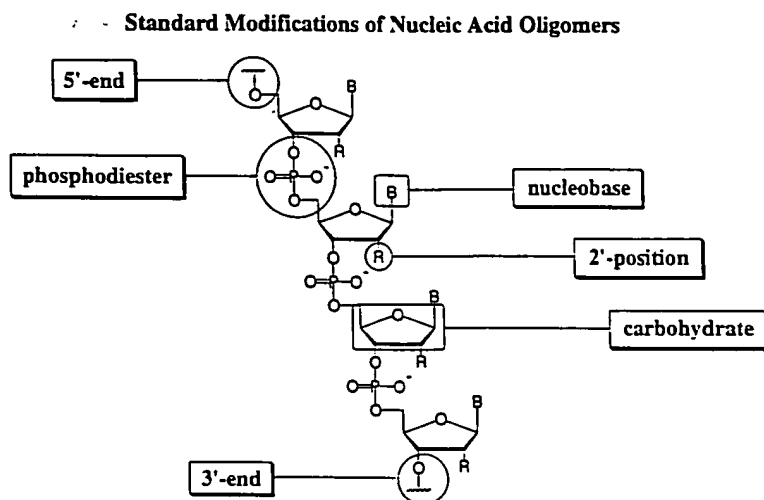


FIG. 12

Autoradiography of PAGE analysis of the ligation of 5-mer 7 with 5'-32 P labeled 12-mer 8. Gel A shows the results of the reaction run with NiCl₂ in imidazole•HCl buffer, while Gel B shows the result of the reaction run with MgCl₂ in MES buffer. Lanes 2 (Gel A) and 1 (Gel B) show the migration of the reaction mixture from ligation on circular DNA template 1. Lanes 1 (Gel A) and 2 (Gel B) compare the results of the ligation reaction on the corresponding single-strand DNA template X. Lane 3 (Gel A) shows the results of the ligation reaction with no template present. Lane 3 (Gel B) shows the migration of the expected full length ligated product which was independently synthesized and 5'-32 P labeled.



Scheme 16



(10) Beaucage, S.L.; Iyer, R.P. *Tetrahedron* 1993, 49, 1925-63.
 (11) Uhlmann, E.; Peyman, A. *Chem Rev.* 1990, 90, 543-84.

Figure 13. Components of nucleic acid polymers that are commonly modified to induce selective properties or functionality to an oligomer.

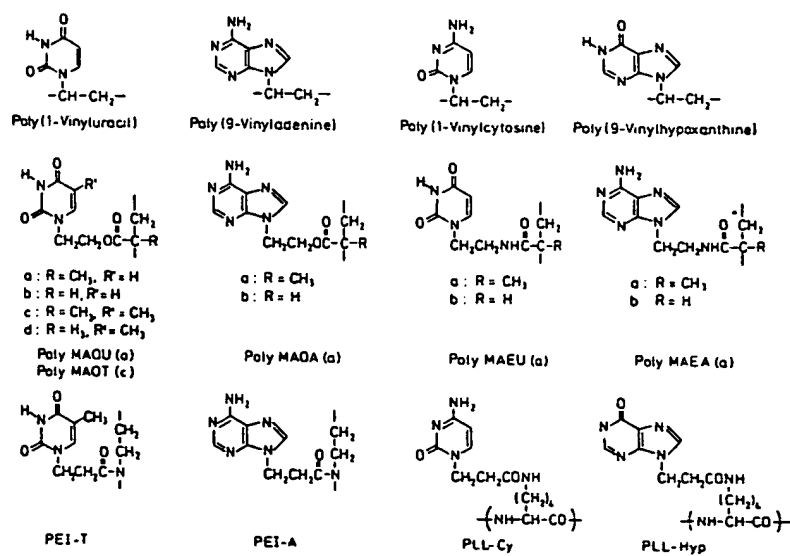


Figure 14. Synthetic Nucleic Acid Analogs.